

A Practical, Highly Stereoselective Synthesis of the Trisaccharide Repeating Unit
of a *Streptococcus pneumoniae* Polysaccharide

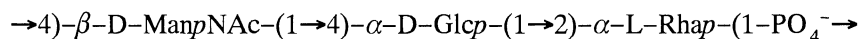
Eisuke KAJI,* Frieder W. LICHTENTHALER,[†] Yumiko OSA, Keiko TAKAHASHI, Eiko MATSUI,
and Shonosuke ZEN

School of Pharmaceutical Sciences, Kitasato University, Minato-ku, Tokyo 108

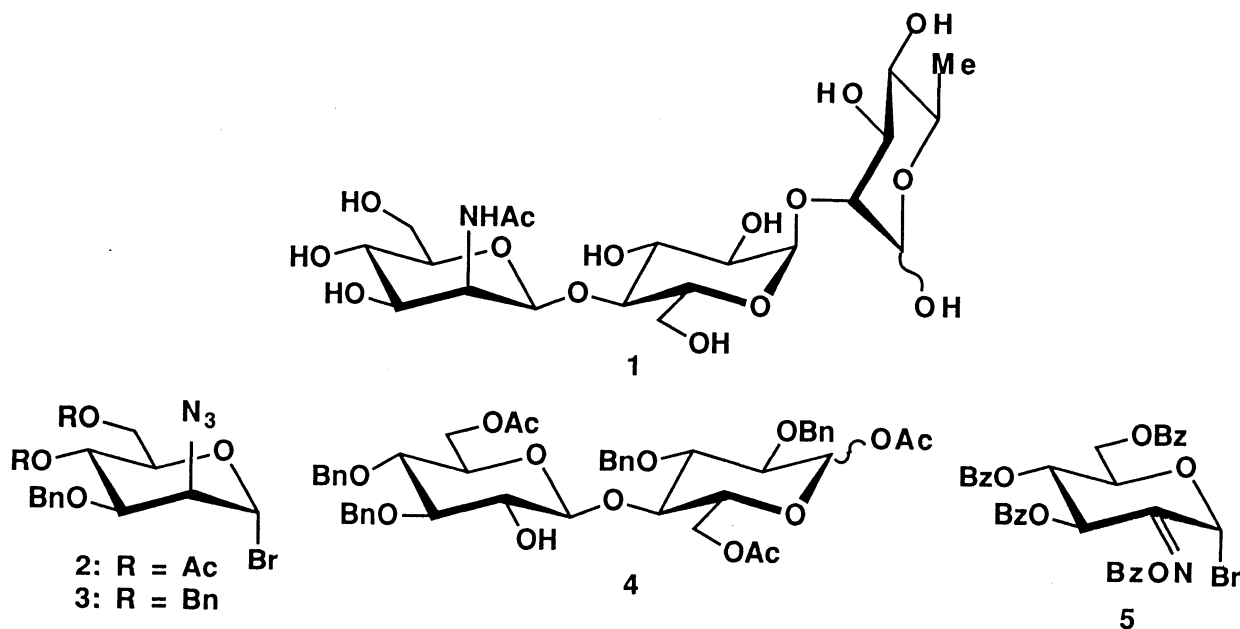
[†]Institut für Organische Chemie, Technische Hochschule Darmstadt, D-6100 Darmstadt, Germany

A highly stereoselective method has been developed for the synthesis of the trisaccharide component of the capsular polysaccharide of *Streptococcus pneumoniae* type 19F. The key intermediate, 2-benzoyloxyiminoglycosyl bromide was successfully utilized for the construction of the *N*-acetyl- β -D-mannosamine unit of the trisaccharide.

Streptococcus pneumoniae type 19F is a major pathogenic bacteria for serious pneumonia,¹⁾ in particular, in infants whose immunity is still less sufficient against diseases. The structure of the repeating unit of the capsular polysaccharide of type 19F bacteria has been elucidated^{2,3)} as *N*-acetyl- β -D-mannosamine-containing trisaccharide:



The trisaccharide component **1** is to be considered an epitope of the antigenic polysaccharide and, hence, an intercellular recognition molecule for the infection. Such biological significance has elicited considerable efforts towards the synthetic acquisition of **1** in quest of artificial antigens, resulting in three different syntheses.⁴⁻⁶⁾ Since direct β -glycosylation of *N*-acetyl-D-mannosamine is not feasible — α -D-manno-



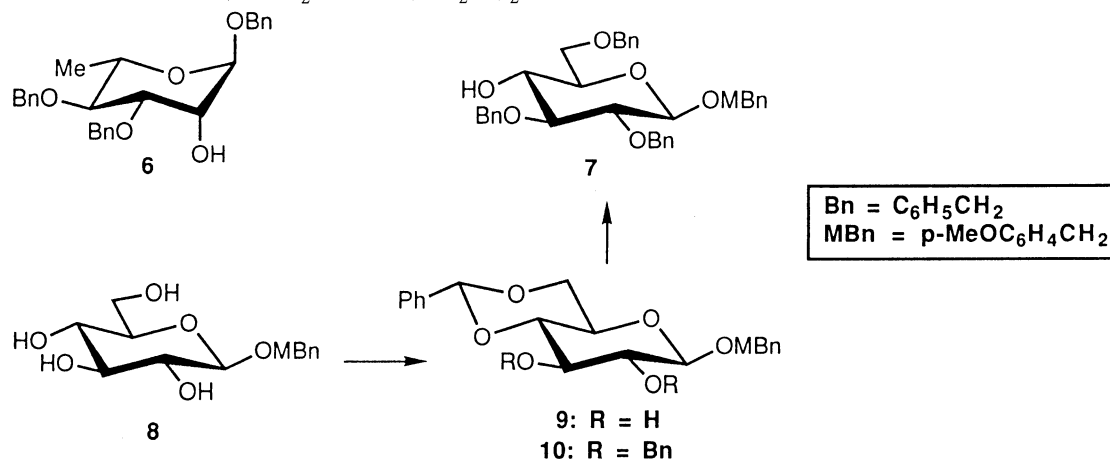
saminides will be formed exclusively —, every approach has employed designed progenitors for the generation of the *N*-acetyl- β -D-mannosamine unit of **1**. Accordingly, 2-azido-2-deoxy-mannopyranosyl halides **2**⁵⁾ and **3**⁶⁾ have been utilized as mannosaminy donors, or alternately, the elaborately blocked cellobiose derivative **4**, generated from its monosaccharide components, in which the equatorial 2'-OH was converted into an axial 2-NHAc via oxidation, oximation, and reduction.⁴⁾ However, neither of these approaches meets practical preparative criteria, inasmuch as they entail more than 16 steps from D-glucose to **1** with overall yields of less than 1%^{4,5)} and 1.8%.⁶⁾

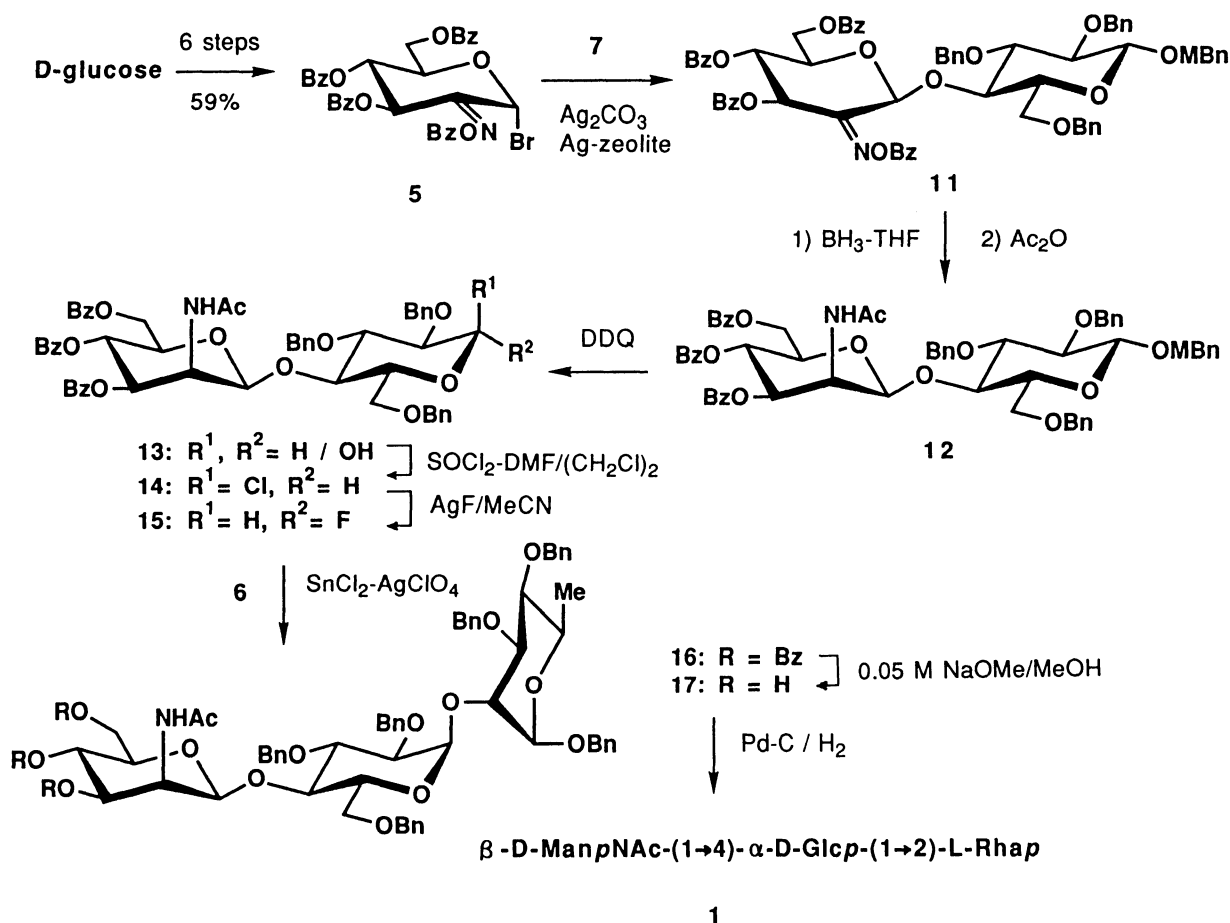
We here describe a novel, preparatively useful synthesis of the target trisaccharide **1** using as the key intermediate, the 2-benzoyloxyiminoglycosyl bromide **5**, which is readily accessible from glucose in 6-high-yielding steps (overall 59%),⁷⁾ and whose utility as a suitable glycosyl donor for the convenient successive elaboration of β -D-mannosamine units has amply been demonstrated.⁸⁾ Given the availability of L-rhamnose derivative **6**⁹⁾ as a suitable glycosyl acceptor for attachment of the rhamnose portion, an appropriately substituted central glucose derivative was required capable of functioning, successively, as a 4-*O*-acceptor and a glycosyl donor. The *p*-methoxybenzyl 2,3,6-tri-*O*-benzyl- β -D-glucoside **7** was selected for this purpose, since, upon glycosylation the anomeric substituent was apt to be removable smoothly by oxidative cleavage with DDQ.¹⁰⁾ The preparation of **7** followed conventional methodology, such that acetobromoglucose was converted into the β -MBn glucoside by reaction with *p*-methoxybenzyl alcohol, followed by de-*O*-acetylation, benzylation (**8**→**9**), benzylation (→**10**), and reductive acetal opening to provide **7** in a 32% overall yield over the 5 steps involved.

The highly β -selective glycosidation of bromide **5** with the 4-OH-free glucoside **7** was effected by using $\text{Ag}_2\text{CO}_3/\text{Ag}$ -zeolite as the catalyst in dichloromethane (48 h, 25 °C), affording an approximate 10:1 (¹H-NMR)¹¹⁾ β/α anomeric mixture of disaccharides from which **11**¹¹⁾ was isolated in 85% yield. If, however, glycosidation of **5** with **7** was induced by silver triflate, the respective α -linked disaccharide¹¹⁾ was obtained in a yield of 90%.

Reduction of the β -disaccharide **11** with borane-THF complex (2 h, 25 °C) proceeded with high preference for hydride attack from the α -side (12:1 based on ¹H-NMR) and subsequent *N*-acetylation with acetic anhydride afforded the *N*-acetyl- β -D-mannosaminy-(1→4)- β -D-glucoside **12**¹⁴⁾ in 69% yield.

In order to convert the disaccharide **12** into a glycosyl donor sufficiently reactive to glycosylate rhamnose **6**, the glycosyl fluoride **15** was generated by way of de-*O*-methoxybenzylation (DDQ/ CH_2Cl_2 - H_2O , →**13**, 43%),¹⁵⁾ chlorination (SOCl_2 -DMF/ $(\text{CH}_2\text{Cl})_2$, →**14**, quant.), and fluorination (AgF/MeCN , 77%).





Indeed, when the fluoride **15** was exposed to the rhamnosyl acceptor **6**⁹⁾ in the presence of $\text{AgClO}_4\text{-SnCl}_2$ in dichloromethane, an essentially α -selective glycosylation was effected to afford the desired trisaccharide **16** as a syrup in a 51% yield. The coupling constants for the central glucosyl moiety ($J_{1,2'} = 3.5$, $J_{2,3'} = 9.0$ Hz) unequivocally proved the correct intersaccharidic link-up. Subsequent de-*O*-benzoylation of **16** with 0.05 M NaOMe/MeOH ($\rightarrow$ **17**, 89%), and hydrogenolysis for removing the benzyl protecting groups (Pd-C/H_2 , MeOH, 94%) proceeded smoothly to the target trisaccharide **1**, which was isolated as an amorphous product of $[\alpha]_{\text{D}}^{22} + 28^\circ$ (c 0.5, MeOH). Its ^1H - and ^{13}C -NMR data¹⁶⁾ were identical in all respects with those reported for the natural²⁾ as well as the synthetic product.^{5,6)}

In summation, a straightforward, highly stereoselective synthesis of the repeating unit of an infectious capsular polysaccharide has been achieved. Our method is not only more efficient — 5% yield over 14 steps from D-glucose — than the previous ones,⁴⁻⁶⁾ but also practical in elaboration of the critical β -D-mannosaminyl portion. The construction of analogs of **1** as well as other, more complex oligosaccharides with a β -D-ManpNAc unit is currently under investigation.

References

- 1) H. J. Jennings, *Adv. Carbohydr. Chem. Biochem.*, **41**, 155 (1983).
- 2) N. Ohno, T. Yadomae, and T. Miyazaki, *Carbohydr. Res.*, **80**, 297 (1980).
- 3) H. J. Jennings, K. G. Rossel, and D. J. Carlo, *Can. J. Chem.*, **58**, 1069 (1980).
- 4) L. Panza, F. Ronchetti, G. Russo, and L. Toma, *J. Chem. Soc., Perkin Trans. 1*, **1987**, 2745.

- 5) T. Sugawara and K. Igarashi, *Carbohydr. Res.*, **172**, 195 (1988).
- 6) H. Paulsen, B. Helpap, and J. P. Lorentzen, *Carbohydr. Res.*, **179**, 173 (1988).
- 7) F. W. Lichtenthaler, P. Jarglis, and W. Hempe, *Liebigs Ann. Chem.*, **1983**, 1959.
- 8) E. Kaji, F. W. Lichtenthaler, T. Nishino, A. Yamane, and S. Zen, *Bull. Chem. Soc. Jpn.*, **61**, 1291 (1988).
- 9) A. Liptak, P. Fugedi, and P. Nanasi, *Carbohydr. Res.*, **65**, 209 (1978).
- 10) Y. Oikawa, T. Yoshida, and O. Yonemitsu, *Tetrahedron Lett.*, **23**, 885 (1982).
- 11) Disaccharide **11** was obtained as a colorless syrup of $[\alpha]_D^{25} + 9.8^\circ$ (c 0.7, CHCl₃), its β -configuration followed from $J_{3,4'}$ and $J_{4,5'}$ couplings of only 5.5 Hz each, indicating distortion of the pyranoid ring towards the twist-boat form as depicted in formula **11**. Similar observations have been made for other β -anomeric 2-benzoyloxyiminoglycosides (Refs. 8, 12, and 13). In contrast, the corresponding α -anomer of **11**, of mp 60–63 °C and $[\alpha]_D^{23} + 43.8^\circ$ (c 0.5, CHCl₃), had the normal $J_{3,4'}$ and $J_{4,5'}$ values of 10 Hz each.
- 12) F. W. Lichtenthaler, E. Kaji, and S. Weprek, *J. Org. Chem.*, **50**, 3505 (1985).
- 13) F. W. Lichtenthaler and E. Kaji, *Liebigs Ann. Chem.*, **1985**, 1659.
- 14) Relevant physical data for **12**: mp 130–132 °C, $[\alpha]_D^{23} - 29.7^\circ$ (c 0.5, CHCl₃); $J_{1,2'} = 1.2$, $J_{2,3'} = 4.0$ Hz.
- 15) Application of another de-*O*-methoxybenzylating agent, ammonium cerium(IV) nitrate was found to be less effective than DDQ.
- 16) The anomeric composition of trisaccharide **1** at the reducing end was about 2:1 in favor of the α -anomer. ¹H-NMR (300 MHz, D₂O): δ = 1.21 (3H, d, α -H-6), 1.22 (3/2H, d, β -H-6), 1.99 (3H, s, NHAc), 3.92 (1/2H, dd, β -H-2), 4.01 (1H, td, α -H-5'), 4.47 (1H, dd, H-2''), 4.81 (1H, d, H-1''), 4.86 (1/2H, broad s, β -H-1), 4.92 (1H, d, α -H-1'), 5.00 (1/2H, d, β -H-1'), 5.14 (1H, d, α -H-1); $J_{1,2} = 1.5$ (α), 1.0 (β), $J_{5,6} = 6.3$ (α), 5.5 (β), $J_{1,2'} = 3.8$ (α), 3.7 (β), $J_{1'',2''} = 1.3$, $J_{2'',3''} = 4.2$ Hz. ¹³C-NMR (75 MHz, D₂O): δ = 17.5 (α - and β -C-6), 22.9 (NHCOMe), 54.1 (C-2''), 60.6 (α - and β -C-6'), 61.2 (C-6''), 67.5 (C-4''), 69.5 (α -C-5), 70.1 (α -C-3), 71.1 (α -C-5'), 71.4 (β -C-5'), 72.0 (α -C-2'), 72.1 (α -C-3'), 72.4 (β -C-4), 72.7 (β -C-2',3'), 72.8 (β -C-3, α -C-4), 72.9 (C-3''), 73.1 (β -C-5), 77.4 (C-5''), 78.2 (α -C-2), 79.3 (β -C-4'), 79.5 (α -C-4'), 81.8 (β -C-2), 92.4 (α -C-1), 94.6 (β -C-1), 98.5 (α -C-1'), 100.2 (C-1''), 101.8 (β -C-1'), 176.3 (NHCO).

(Received December 19, 1991)